LB Plates and Liquid
Thursday, November 03, 2011
2:49 PM
Sean Weise

LB Plates, 200 ml
In 200 ml dH₂O
+ 5 g LB Broth (Difco, Luria-Bertani)
+ 3 g Agar (Fisher, BP 1423-500)
Autoclave for 20 min, swirl immediately, allow to cool to touch
Pour plates (~ 35 ml) label, store at 4°C

LB Liquid, 200 ml
In 200 ml dH₂O
+ 5 g LB Broth (Difco, Luria-Bertani)
Autoclave for 20 min

<table>
<thead>
<tr>
<th>Antibiotic</th>
<th>Stock (mg/ml)</th>
<th>Working (µg/ml)</th>
<th>µl of stock in 5 ml</th>
<th>µl of stock in 20 ml</th>
<th>µl of stock in 40 ml</th>
<th>ml of stock in 200 ml</th>
<th>ml of stock in 500 ml</th>
<th>ml of stock in 1 L</th>
</tr>
</thead>
<tbody>
<tr>
<td>Ampicillin</td>
<td>10</td>
<td>50</td>
<td>25</td>
<td>100</td>
<td>200</td>
<td>1</td>
<td>2.5</td>
<td>5</td>
</tr>
<tr>
<td>Chloramphenicol (methanol)</td>
<td>10</td>
<td>20</td>
<td>10</td>
<td>40</td>
<td>80</td>
<td>0.4</td>
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<td>2</td>
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<tr>
<td>Kanamycin</td>
<td>10</td>
<td>25</td>
<td>12.5</td>
<td>50</td>
<td>100</td>
<td>0.5</td>
<td>1.25</td>
<td>2.5</td>
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<tr>
<td>Rifampicin (methanol)</td>
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<td>150</td>
<td>25</td>
<td>100</td>
<td>200</td>
<td>1</td>
<td>2.5</td>
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<tr>
<td>Spectinomycin</td>
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<td>100</td>
<td>50</td>
<td>200</td>
<td>400</td>
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<td>10</td>
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<tr>
<td>Streptomycin</td>
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<td>30</td>
<td>15</td>
<td>60</td>
<td>120</td>
<td>0.6</td>
<td>1.5</td>
<td>3</td>
</tr>
</tbody>
</table>

Note: If using multiple antibiotics you can cut the concentration of each by half

10 mg/ml Antibiotic, 10 ml
In 10 ml of sterile dH₂O
+ 0.1g Antibiotic
Filter sterilize using 0.2 - 0.45 µm syringe filter
Aliquot into sterile 1.5 ml microfuge tubes and store at -20°C

Glycerol Stock
1. To make glycerol stock prepare a sterile cryovile by adding 800 µl of 80% sterile glycerol
2. To the cryovile add 1 ml of E.Coli culture
3. Snap freeze in LN₂
Note: Try to make Glycerol stocks from E. Coli in the log phase of growth. i.e. OD = 2